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# A novel fluorescein derivative as a colorimetric chemosensor for detecting copper(II) ion

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#### ABSTRACT

A novel fluorescein derivative, synthesized by the reaction of fluorescein hydrazide and 1-phenyl-3-methyl-4-benzoyl-5-pyrazolone, was evaluated as a chemoselective metal ion sensor. Addition of  $Cu^{2+}$  to an aqueous solution of the fluorescein derivative resulted in a rapid color change from colorless to deep yellow together with a distinctive change in UV–vis absorption spectrum. However, other common alkali-, alkaline earth-, transition- and rare earth metal ions induced no or minimal spectral changes. The stoichiometry of the reaction and association constant of the fluorescein derivative with  $Cu^{2+}$  are described. Experimental results indicate that the fluorescein derivative could provide a rapid, selective and sensitive response to  $Cu^{2+}$ , and could be used as a potential  $Cu^{2+}$  colorimetric chemosensor in aqueous solution.

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#### 1. Introduction

Metal ion sensors are employed in applications ranging from clinical toxicology, environmental bioinorganic chemistry, bioremediation, and waste management [1–3] and much attention has focussed to the development of sensing devices for copper(II). As the third most abundant divalent metal ion in the human body, after Fe<sup>2+</sup> and Zn<sup>2+</sup>, Cu<sup>2+</sup> plays a pivotal role in a variety of fundamental physiological processes in organisms ranging from bacteria to mammals [4,5]. Copper compounds are also employed for plant diseases treatment, water treatment and as preservatives for wood and leather. Nonetheless, while a low-level background intake of copper is indispensable, high doses of copper can be harmful and even toxic to biological systems [4,6].

Many sensing methods for detecting  $Cu^{2+}$  have been described, such as colorimetric and fluorescent chemosensors, and electrochemical methods [7–9]. Colorimetric sensors are promising due to the simplicity of the assay. Furthermore, colorimetric assays have a significantly lower capital cost than closely related methods, such

as fluorescent sensors, for which both spectrophotometric equipment and a UV light source are required [10–16].

The fluorescein and rhodamine family of dyes, with spirolactam structure (closed form) are non-fluorescent. As shown in Fig. 1, ring-opening of the spirolactam gives the open form and results in a strong fluorescence and obvious color change [17–20]. Due to large visible-range extinction coefficients and high fluorescence quantum yields for fluorescein and rhodamine, these compounds are excellent antenna chromophores [21–26].

In recent years, several rhodamine-based chemosensors and chemodosimeters for metal ions, such as Cu<sup>2+</sup> [27–30], Hg<sup>2+</sup> [20,31–34], Fe<sup>3+</sup> [35], and Pb<sup>2+</sup> [19] have been studied. The cationsensing mechanism of these probes is based on the change in structure between spirocyclic and opencyclic forms. However, fluorescein-based probes have received comparatively little attention [36,37]. We synthesized a fluorescein-based colorimetric chemosensor, 1-phenyl-3-methyl-5-hydroxypyrazole-4-benzoyl (fluorescein)hydrazone (1), for rapid, selective and sensitive response to Cu<sup>2+</sup> in aqueous media (Fig. 2). Solutions of 1 are colorless, but upon addition of micromolar Cu<sup>2+</sup> a deep yellow color is obtained. Addition of other common alkali-, alkaline earth, transition- and rare earth metal ions result in no or minimal spectral change. Compound 1 is a naked-eye chemosensor for

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Fig. 1. Structure of fluorescein, rhodamine B and rhodamine 6G.

detection of  $Cu^{2+}$  that upon chelation of  $Cu^{2+}$  1 will change to a strongly colored ring-opened form.

#### 2. Experimental

#### 2.1. Materials and instruments

All the materials for synthesis were purchased from commercial suppliers and used without further purification. With the exception of  $Al(NO_3)_3$  and  $MgSO_4$ , aqueous solutions of metal ions were prepared from their chloride salts.

NMR spectra were taken on a Bruker Avance DRX-200 or Varian Mercury Plus-300 BB spectrometer with TMS (tetramethylsilane) as internal standard and DMSO- $d_6$  as solvent. Mass spectra were obtained on a Bruker Esquire 6000 spectrometer. UV—vis absorption spectra were obtained with a Perkin Elmer Lambda 35 UV—vis spectrophotometer and recorded in quartz cells with 1 cm optical path length. Fluorescence spectra were recorded on a Hitachi RF-4500 fluorescence spectrophotometer.

#### 2.2. General procedures of metal ion sensing

#### 2.2.1. Job's plot analyses

For Job's plot analyses (in Section 3.3), a  $1.0\times10^{-3}$  M stock solution of  $Cu^{2+}$  in  $H_2O$ , and  $1.0\times10^{-3}$  M stock solution of 1 in DMSO (dimethylsulfoxide) were prepared. The  $Cu^{2+}$  stock solution and the stock solution of 1 were placed in a test tube to obtain a total volume of 20  $\mu$ L, then diluted to 2 mL with DMSO/ $H_2O$  (4:6, v/v) to get the test solution (nine test solutions were got; for the

Fig. 2. Synthetic route of chemosensor 1.

first test solution, 2  $\mu$ L stock solution of  $Cu^{2+}$  and 18  $\mu$ L stock solution of **1** were used, then diluted to 2 mL with DMSO/H<sub>2</sub>O (4:6, v/v) to get the first test solution; for the second test solution, 4  $\mu$ L stock solution of  $Cu^{2+}$  and 16  $\mu$ L stock solution of **1** were used; for the third test solution, 6  $\mu$ L stock solution of  $Cu^{2+}$  and 14  $\mu$ L stock solution of **1** were used; until 18  $\mu$ L stock solution of  $Cu^{2+}$  and 2  $\mu$ L stock solution of **1** were used to get the ninth test solution). All the analyses were performed with  $[1] + [Cu^{2+}] = 10.0 \ \mu$ M. The absorbance at 495 nm of these test solutions was recorded.

#### 2.2.2. Spectral analyses

In other spectral analyses, a  $1.0 \times 10^{-3}$  M solution of **1** was prepared in DMSO, and then diluted by DMSO/H<sub>2</sub>O (4:6, v/v) to give a  $1.0 \times 10^{-5}$  M stock solution of **1** for spectral analyses. Stock solutions of metal ions were prepared in H<sub>2</sub>O ( $2.0 \times 10^{-3}$  M of Cu<sup>2+</sup> was used in Section 3.2;  $2.0 \times 10^{-2}$  M of metal ions were used in other sections, except Sections 3.2 and 3.3). Each time, a 2 ml stock solution of **1** ( $1.0 \times 10^{-5}$  M) was added to the quartz cell, and the required quantity of stock solution of metal ions was added with a microsyringe.

All the measurements were taken at room temperature about 298 K. After 2 min of the mixing of metal ions with 1, UV—vis absorption spectra or fluorescence emission spectra were measured, unless otherwise indicated. For all fluorescent tests, excitation wavelength was 287 nm, with excitation and emission slit widths both 5 nm. Addition of metal ions increased the volume by no more than 0.04 mL, so that dilution was insignificant.

#### 2.3. Synthesis

#### 2.3.1. Synthesis of fluorescein hydrazide (2)

In a 100 mL flask containing a suspension of fluorescein (6 g, 18.1 mmol) in 50 mL methanol, excess hydrazine hydrate (24 mL; hydrazine content >80 mass%) was added. The reaction mixture was heated to reflux for 7 h with stirring, during which time the suspended particles were consumed and a clear solution was obtained. The ensuing solution was allowed to cool and poured into 400 mL H<sub>2</sub>O at which time, a yellow precipitated formed immediately, which was allowed to settle for 2 h. The aqueous suspension was filtered, washed with water until the filtrate was colorless, and washed 3  $\times$  10 mL with cold absolute ethanol. The crude product was purified by recrystallization from ethanol to give  $3.59 \text{ g of } \mathbf{2}$  as an off-white solid (57%). Melting point:  $262-264 \,^{\circ}\text{C}$ . <sup>1</sup>H NMR (200 MHz, DMSO- $d_6$ ),  $\delta$  (ppm): 4.38 (s, 2H), 6.42 (m, 4H), 6.59 (s, 2H), 6.97 (m, 1H), 7.48 (m, 2H), 7.76 (m, 1H), 9.81 (s, 2H). <sup>13</sup>C NMR (50 MHz, DMSO- $d_6$ ),  $\delta$  (ppm): 64.6, 102.3, 109.9, 111.9, 122.3, 127.9, 128.4, 129.3, 132.6, 151.5, 152.4, 158.1, 165.4.

## 2.3.2. Synthesis of 1-phenyl-3-methyl-5-hydroxypyrazole-4-benzoyl(fluorescein)hydrazone (1)

In a 25 mL flask, **2** (0.346 g, 1 mmol) and 1-phenyl-3-methyl-4-benzoyl-5-pyrazolone (PMBP; 0.278 g, 1 mmol) were suspended in 10 mL methanol. The mixture was refluxed for 8 h with stirring, during which time a clear solution formed. Following reaction, the mixture was allowed to cool to room temperature and a yellow precipitate formed. The precipitated was separated by filtration and washed with  $3 \times 10$  mL methanol. After drying, 0.25 g of **1** (bright yellow solid) was prepared in 41% yield. Melting point >320 °C. <sup>1</sup>H NMR (300 MHz, DMSO- $d_6$ ),  $\delta$  (ppm): 1.23 (s, 3H), 5.96 (s, 2H), 6.35 (s, 2H), 6.61 (d, J = 2.1 Hz, 3H), 7.12 (m, 3H), 7.21 (s, 2H), 7.36 (m, 3H), 7.63 (m, 2H), 7.77 (d, J = 8.1 Hz, 2H), 7.89 (d, J = 6.9 Hz, 1H), 10.04 (s, 2H), 11.51 (s, 1H). <sup>13</sup>C NMR (75 MHz, DMSO- $d_6$ ),  $\delta$  (ppm): 15.1, 66.4, 100.7, 102.4, 107.5, 113.2, 118.2, 123.3, 124.4, 127.1, 127.7, 128.7, 129.5, 130.2, 134.7, 138.3, 147.4, 149.6, 152.8, 159.0, 164.7

**Fig. 3.** According to literature, the reaction mechanism of **2** with Cu<sup>2+</sup>.

166.7, 167.3. ESI mass spectrometry, m/z: 607.3 (M + H)<sup>+</sup>, 629.3 (M + Na)<sup>+</sup>.

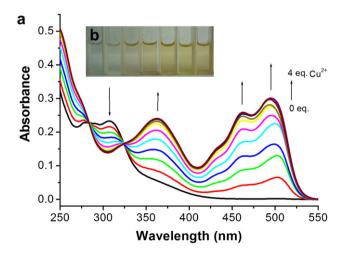
#### 3. Results and discussion

#### 3.1. Design and synthesis of sensor 1

In past few years, some fluorescein-based metal ions sensors were reported. Lippard and co-workers installed a series of one or two binding units, such as 8-aminoquinoline, methylated dipicolylamine, or thioether on the xanthene moiety of fluorescein [38–42]. Similar work was also reported by Jun et al. [43] and Kim et al. [44]. Guo and co-workers attached some chelator groups to the benzene moiety of decarboxylated fluorescein [45,46].

However, fluorescein-based metal ions sensors, which are based on the change in structure between spirocyclic and opencyclic forms of fluorescein moiety, have been seldom investigated up to now [36,37]. According to Ma and co-workers, the parent compound **2** can be used as a selective and sensitive probe for  $Cu^{2+}$  [36]. In aqueous solution,  $Cu^{2+}$  can be recognized and bound by compound **2**, and the subsequent complexation of  $Cu^{2+}$  promotes hydrolytic cleavage of the amide bond, causing the release of fluorophore (fluorescein) and thereby the retrievement of fluorescence (Fig. 3). A reaction time of 2 h at room temperature was used for their detecting system. Thus, compound **2** can not be used for real-time monitoring of  $Cu^{2+}$ .

Our considerations for synthesizing **1** are as follows: (a) PMBP is known to possess a high chelating ability with common metal ions [47–49]. Introducing PMBP moiety can generate the chelating ability of **1** toward some metal ions; (b) when coordination takes place, the spirolactam of fluorescein moiety can be opened, at the



**Fig. 4.** UV—vis absorption spectra (a) and photographs of color changes (b) of **1** (10.0  $\mu$ M) with gradual addition of Cu<sup>2+</sup> [(a) 0, 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, and 40  $\mu$ M, respectively; (b) from left to right: 0, 2, 6, 10, 16, 20, and 40  $\mu$ M, respectively] in aqueous solution (DMSO/H<sub>2</sub>O = 4:6, v/v).

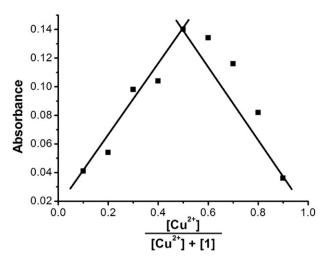


Fig. 5. Job's plot of 1 with  $Cu^{2+}$  in aqueous solution (DMSO/H $_2$ O = 4:6, v/v). Total concentration of 1 +  $Cu^{2+}$  was kept constant at 10.0  $\mu$ M. The absorbance at 495 nm was used.

same time obvious color and optical change can be observed. Consequently, sensor **1** was synthesized via a simple two-step reaction, and confirmed by melting point, <sup>1</sup>H NMR, <sup>13</sup>C NMR, and mass spectrum.

#### 3.2. UV-vis titration of **1** with $Cu^{2+}$

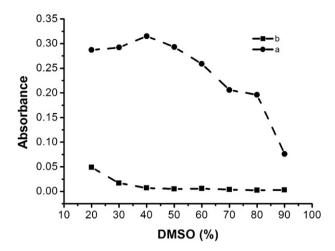
Fig. 4a shows the UV—vis absorption spectra of **1** with various concentrations of  $\text{Cu}^{2+}$  in aqueous solution at room temperature. In aqueous solution (DMSO/H<sub>2</sub>O = **4**:6, v/v), **1** (10.0  $\mu$ M) had nearly no absorption in the visible region, which was ascribed to the closed spirolactam form of **1**. The characteristic peak of 66.4 ppm (9-carbon) in <sup>13</sup>C NMR spectrum of **1** also supported this dentification.

Each time, 2  $\mu$ L of Cu<sup>2+</sup> stock solution (2.0  $\times$  10<sup>-3</sup> M) was added to the stock solution of 1 (10.0  $\mu$ M). For the spectra of 1, upon addition of Cu<sup>2+</sup>, the band at 307 nm decreased and three new bands at 363 nm, 460 nm, and 495 nm appeared, respectively. Correspondingly, the solution of 1 changed from colorless to deep yellow (Fig. 4b). The significant color change indicates that 1 is a sensitive naked-eye indicator for Cu<sup>2+</sup>.

According to linear Benesi–Hildebrand expression, the measured absorbance  $[1/(A-A_0)]$  at 370 nm varied as a function of  $1/[Cu^{2+}]$  in a linear relationship (R=0.9932), indicating formation of 1:1 stoichiometry between  $Cu^{2+}$  and 1 [13,50,51]. The association constant of 1 with  $Cu^{2+}$  in the solution of DMSO/H<sub>2</sub>O (4:6, v/v) was calculated to be  $3.3 \times 10^4$  M<sup>-1</sup>.

Reactions of 1 with  $Cu(AcO)_2$ ,  $CuSO_4$ , or  $Cu(NO_3)_2$  gave the same changes as the cases of 1-CuCl<sub>2</sub> on the UV—vis absorption spectra, demonstrating that counter-anion effect is negligible.

Fig. 6. Proposed binding mechanism of 1 with Cu<sup>2+</sup>.



**Fig. 7.** Effect of DMSO content on the absorbance of **1** (10.0  $\mu$ M) at 495 nm in the presence (a) and absence (b) of  $Cu^{2+}$  (20.0  $\mu$ M) in DMSO aqueous solution.

#### 3.3. Job's plot analyses

For further determination the stoichiometry between 1 and  $Cu^{2+}$ , Job's plot analyses were also used. Total concentration of  $1 + Cu^{2+}$  was kept constant at 10.0  $\mu$ M. When molar fraction of  $Cu^{2+}$  was 0.5, the absorbance at 495 nm got to maximum (Fig. 5), indicating that forming a 1:1 complex between 1 and  $Cu^{2+}$ .

#### 3.4. Binding mode studies

Usually, for most of rhodamine-based probes and a few fluorescein-based probes, the cation-sensing mechanism is based on the change in structure between spirocyclic and opencyclic forms. In the absence of metal ions, these probes exist in the spirocyclic forms which are colorless and non-fluorescent. Addition of metal ions makes the spirocycle open via coordination or irreversible chemical reaction, resulting in color change and generation of strong fluorescence [27–37].

For sensor 1, obvious color and electronic absorption spectrum changes appeared with the addition of  $Cu^{2+}$ . The possible reason is that sensor 1 coordinates to  $Cu^{2+}$ , and meanwhile spirocyclic opening in fluorescein backbone happens (Fig. 6). As a result of the coordination, the absorption bands at 307 nm decrease and the

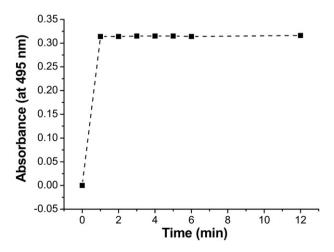
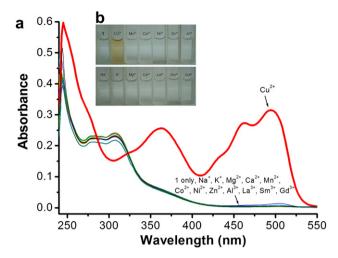


Fig. 8. Time course of the response of 1 (10.0  $\mu M$ ) to 2 equiv. of  $Cu^{2+}$  in aqueous solution (DMSO/H<sub>2</sub>O = 4:6, v/v).



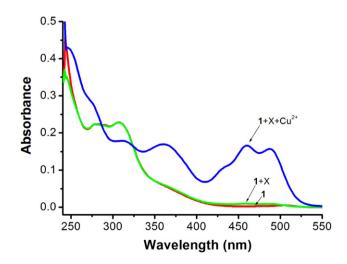
**Fig. 9.** UV–vis absorption spectra (a) and photographs of color changes (b) of **1** (10.0  $\mu$ M) upon addition of various metal ions (20.0  $\mu$ M) in aqueous solution (DMSO/H<sub>2</sub>O = 4:6.  $\nu$ V).

absorption bands at 363 nm increase simultaneously. Isosbestic points at 284 nm and 324 nm also indicate the presence of a new metal complex in equilibrium with the free ligand 1 [16]. Meanwhile, the spirocyclic opening in fluorescein backbone gives the characteristic absorption of fluorescein at 460 nm and 495 nm in the visible region [52]. They are responsible for the color change from colorless to deep yellow.

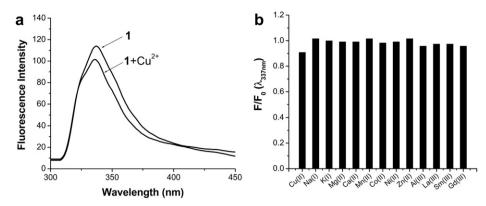
On addition of EDTANa<sub>2</sub> (Ethylenediaminetetraacetic acid disodium salt) to the solution of  $\bf 1$  containing  $Cu^{2+}$ , the solution changed from deep yellow to colorless, indicating a reversible coordination between  $\bf 1$  and  $Cu^{2+}$ . The reversible response toward  $Cu^{2+}$  implies that  $\bf 1$  is a chemosensor not a chemodosimeter of  $Cu^{2+}$ .

#### 3.5. Effect of reaction media and reaction time

For practical applicability, the  $Cu^{2+}$  detection were carried out in aqueous solution (DMSO mixed with water). In order to get optimal experimental phenomenon, the effect of DMSO content on the absorbance of 1 was studied.



**Fig. 10.** UV—vis absorption spectra of **1** (10.0  $\mu$ M) in aqueous solution (DMSO/  $H_2O=4:6$ , v/v) in the presence of  $Cu^{2+}$  (20.0  $\mu$ M) and miscellaneous cations (X) including Na<sup>+</sup>, K<sup>+</sup>, Mg<sup>2+</sup>, Ca<sup>2+</sup>, Mn<sup>2+</sup>, Co<sup>2+</sup>, Ni<sup>2+</sup>, Zn<sup>2+</sup>, Al<sup>3+</sup>, La<sup>3+</sup>, Sm<sup>3+</sup>, and Gd<sup>3+</sup> (20.0  $\mu$ M, respectively).



**Fig. 11.** (a) Fluorescence emission spectra of **1** (10.0  $\mu$ M) and that after addition of Cu<sup>2+</sup> (20.0  $\mu$ M) in aqueous solution (DMSO/H<sub>2</sub>O = 4:6, v/v). (b) Changes in fluorescence emission intensity ratio (F/F<sub>0</sub>) at 337 nm of **1** (10.0  $\mu$ M) in the presence of various metal ions in aqueous solution (DMSO/H<sub>2</sub>O = 4:6, v/v). F<sub>0</sub> is the intensity of free **1**, and F is the intensity of **1** after addition of metal ion. Excitation wavelength was 287 nm.

As shown in Fig. 7, for the free **1**, when DMSO concentration is at or above 40%, **1** has very weak absorbance at 495 nm, therefore, gives lower background signal. In the presence of 2 equiv. of  $Cu^{2+}$ , when DMSO concentration is above 40%, the absorbance of **1** (at 495 nm) dramatically decreased. It means that when DMSO concentration is at 40%, the detection system can give lower background signal and maximum detection signal. As a result,DMSO/ $H_2O = 4:6$  (volume ratio) as the solvent was employed in all spectral analysis experiments.

The effect of reaction time on the UV—vis absorption spectra of the detecting system was studied and the results are shown in Fig. 8. Adding 2 equiv. of  $\text{Cu}^{2+}$  to aqueous solution of **1**, 1 min later the absorbance at 495 nm of the system gets to maximum and does not increase with more reaction time. It means the spirocycle of **1** can be opened in very short time, and **1** could be used as a real-time  $\text{Cu}^{2+}$  chemosensor. 2 min was selected as the detection time in this paper.

#### 3.6. Interference from other metal ions

Selectivity is an important characteristic of an ion-selective chemosensor. To examine the selectivity of **1**, the affinity of **1** for other surveyed metal ions, such as Na<sup>+</sup>, K<sup>+</sup>, Mg<sup>2+</sup>, Ca<sup>2+</sup>, Mn<sup>2+</sup>, Co<sup>2+</sup>, Ni<sup>2+</sup>, Zn<sup>2+</sup>, Al<sup>3+</sup>, La<sup>3+</sup>, Sm<sup>3+</sup> and Gd<sup>3+</sup>, was investigated. As shown in Fig. 9, addition 2 equiv. of these alkali-, alkaline earth-, transition- and rare earth metal ions do not cause significant UV–vis absorption spectral and color changes of **1**. However, under the same conditions, addition of Cu<sup>2+</sup> results in an obvious change, which indicates that chemosensor **1** has higher binding affinity toward Cu<sup>2+</sup> than other surveyed metal ions.

Furthermore, to explore the utility of **1** as a cation-selective chromogenic chemosensor, competition experiment was carried out by adding 20.0  $\mu$ M Cu²+ to solution of **1** (10.0  $\mu$ M) in the presence of miscellaneous cations including Na+, K+, Mg²+, Ca²+, Mn²+, Co²+, Ni²+, Zn²+, Al³+, La³+, Sm³+, and Gd³+ (20.0  $\mu$ M, respectively; Fig. 10). These miscellaneous competitive cations did not lead to any significant absorption changes of **1**. However, upon addition of 2 equiv. of Cu²+ to the solution, the unique spectral and color changes were still displayed. These results indicate that **1** has a good selectivity toward Cu²+ over other competitive cations, and more, the detection of Cu²+ by **1** is hardly affected by these common coexistent metal ions.

#### 3.7. Influence of metal ions on the fluorescence intensity of 1

The fluorescence titration of 1 (10.0  $\mu$ M) with 2 equiv. of metal ions was conducted in aqueous solution (DMSO/H<sub>2</sub>O = 4:6, v/v).

Excitation wavelength was 287 nm, and fluorescence emission intensity was recorded at 337 nm.

In aqueous solution, the fluorescence intensity of **1** was extremely weak. It was also due to the closed spirolactam form of **1**. As shown in Fig. 11, upon addition of 2 equiv. of Cu<sup>2+</sup>, emission intensity of **1** was slightly quenched. Addition of 2 equiv. of other surveyed metal ions (Na<sup>+</sup>, K<sup>+</sup>, Mg<sup>2+</sup>, Ca<sup>2+</sup>, Mn<sup>2+</sup>, Co<sup>2+</sup>, Ni<sup>2+</sup>, Zn<sup>2+</sup>, Al<sup>3+</sup>, La<sup>3+</sup>, Sm<sup>3+</sup> and Gd<sup>3+</sup>, respectively) almost did not produce a change in the fluorescence spectra of **1**, in terms of maximum emission intensity and general peak shape. Fluorescence intensity ratios of **1** in the presence and absence of these surveyed metal ions at 337 nm are almost equal to one (Fig. 11b).

After coordination to  $Cu^{2+}$ , **1** only gave the characteristic color change, and did not generate the strong fluorescence emission of fluorescein moiety. We attribute this to intramolecular fluorescence quenching by paramagnetic  $Cu^{2+}$ . Similar phenomena was also observed in other experiments [44,53].

Quenching of the fluorescence of luminescent ligand by coordination to a paramagnetic Cu<sup>2+</sup> center can be restored by nitric oxide-induced reduction to a diamagnetic Cu<sup>+</sup> species. It is a strategy for metal-based nitric oxide sensing [54]. Can **1-**Cu<sup>2+</sup> complex be used as a nitric oxide sensor? It deserves further research.

#### 4. Conclusions

In conclusion, a novel fluorescein derivative (1) has been synthesized by the reaction of fluorescein hydrazide and 1-phenyl-3-methyl-4-benzoyl-5-pyrazolone. Addition of  $Cu^{2+}$  to an aqueous solution (DMSO/H $_2$ O = 4:6, v/v) of 1 can result in an obvious color change (from colorless to deep yellow) and UV—vis absorption spectral change in a very short time. This makes the real-time, simple-to-use and naked-eye detection of  $Cu^{2+}$  possible. As a result of coordination of 1 with  $Cu^{2+}$ , the spirocyclic opening of fluorescein moiety in 1 is proposed. Moreover, 1 possesses a good selectivity toward  $Cu^{2+}$  over other competitive cations (Na $^+$ , K $^+$ , Mg $^{2+}$ ,  $Ca^{2+}$ , Mn $^{2+}$ ,  $Co^{2+}$ , Ni $^{2+}$ ,  $Zn^{2+}$ , Al $^{3+}$ , La $^{3+}$ , Sm $^{3+}$  and  $Cu^{3+}$ ). These features indicate that 1 is a good candidate, for rapid, selective and sensitive detecting  $Cu^{2+}$  in aqueous media.

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#### References

- de Silva AP, Gunaratne HQN, Gunnlaugsson T, Huxley AJM, McCoy CP, Rademacher JT, et al. Signaling recognition events with fluorescent sensors and switches. Chemical Reviews 1997;97(5):1515–66.
- [2] de Silva AP, Fox DB, Huxley AJM, Moody TS. Combining luminescence, coordination and electron transfer for signalling purposes. Coordination Chemistry Reviews 2000;205(1):41–57.
- [3] Valeur B, Leray I. Design principles of fluorescent molecular sensors for cation recognition. Coordination Chemistry Reviews 2000;205(1):3–40.
- [4] Martínez R, Espinosa A, Tárraga A, Molina P. Bis(indolyl)methane derivatives as highly selective colourimetric and ratiometric fluorescent molecular chemosensors for Cu<sup>2+</sup> cations. Tetrahedron 2008;64(9):2184–91.
- [5] Kaur S, Kumar S. Photoactive chemosensors 4: a Cu<sup>2+</sup> protein cavity mimicking fluorescent chemosensor for selective Cu<sup>2+</sup> recognition. Tetrahedron Letters 2004;45(26):5081-5.
- [6] Aksuner N, Henden E, Yilmaz I, Cukurovali A. Selective optical sensing of copper(II) ions based on a novel cyclobutane-substituted Schiff base ligand embedded in polymer films. Sensors and Actuators B 2008;134(2):510-5.
- [7] Tan J, Yan XP. 2,1,3-Benzoxadiazole-based selective chromogenic chemosensor for rapid naked-eye detection of Hg<sup>2+</sup> and Cu<sup>2+</sup>. Talanta 2008;76 (1):9–14.
- [8] Dujols V, Ford F, Czarnik AW. A long-wavelength fluorescent chemodosimeter selective for Cu(II) ion in water. Journal of the American Chemical Society 1997;119(31):7386-7.
- [9] Singh LP, Bhatnagar JM. Copper(II) selective electrochemical sensor based on Schiff base complexes. Talanta 2004;64(2):313–9.
- [10] Yoosaf K, Ipe BJ, Suresh CH, Thomas KG. In situ synthesis of metal nanoparticles and selective naked-eye detection of lead ions from aqueous media. The Journal of Physical Chemistry C 2007;111(34):12839–47.
- [11] Gunnlaugsson T, Leonard JP, Murray NS. Highly selective colorimetric nakedeye Cu(II) detection using an azobenzene chemosensor. Organic Letters 2004;6(10):1557–60.
- [12] Lee MH, Cho BK, Yoon J, Kim JS. Selectively chemodosimetric detection of Hg (II) in aqueous media. Organic Letters 2007;9(22):4515–8.
- [13] Zhu M, Yuan M, Liu X, Xu J, Lv J, Huang C, et al. Visible near-infrared chemosensor for mercury ion. Organic Letters 2008;10(7):1481–4.
- [14] Lee SJ, Lee SS, Jeong IY, Lee JY, Jung JH. Azobenzene coupled chromogenic receptors for the selective detection of copper(II) and its application as a chemosensor kit. Tetrahedron Letters 2007;48(3):393–6.
- [15] Sheng R, Wang P, Liu W, Wu X, Wu S. A new colorimetric chemosensor for Hg<sup>2+</sup> based on coumarin azine derivative. Sensors and Actuators B 2008;128 (2):507-11.
- [16] Mu H, Gong R, Ma Q, Sun Y, Fu E. A novel colorimetric and fluorescent chemosensor: synthesis and selective detection for Cu<sup>2+</sup> and Hg<sup>2+</sup>. Tetrahedron Letters 2007;48(31):5525-9.
- [17] Maciej A, Jonathan G. Synthesis of novel spirolactams by reaction of fluorescein methyl ester with amines. Tetrahedron Letters 2000;41(6):807–9.
- [18] Maciej A, Jonathan G. Efficient synthesis of rhodamine conjugates through the 2'- position. Bioorganic & Medicinal Chemistry Letters 2000;10(14):1539–41.
- [19] Kwon JY, Jang YJ, Lee YJ, Kim KM, Seo MS, Nam W, et al. A highly selective fluorescent chemosensor for Pb<sup>2+</sup>. Journal of the American Chemical Society 2005;127(28):10107–11.
- [20] Wu JS, Hwang IC, Kim KS, Kim JS. Rhodamine-based Hg<sup>2+</sup>-selective chemodosimeter in aqueous solution: fluorescent off-on. Organic Letters 2007;9 (5):907-10.
- [21] Zumbuehl A, Jeannerat D, Martin SE, Sohrmann M, Stano P, et al. An amphotericin B-fluorescein conjugate as a powerful probe for biochemical studies of the membrane. Angewandte Chemie International Edition 2004;43 (39):5181–5.
- [22] Han J, Jose J, Mei E, Burgess K. Chemiluminescent energy-transfer cassettes based on fluorescein and nile red. Angewandte Chemie International Edition 2007;46(10):1684-7.
- [23] Park J, Lee HY, Cho MH, Park SB. Development of a Cy3-labeled glucose bioprobe and its application in bioimaging and screening for anticancer agents. Angewandte Chemie International Edition 2007;46(12):2018–22.
- [24] Chang PV, Prescher JA, Hangauer MJ, Bertozzi CR. Imaging cell surface glycans with bioorthogonal chemical reporters. Journal of the American Chemical Society 2007;129(27):8400–1.
- [25] Nakayama-Ratchford N, Bangsaruntip S, Sun X, Welsher K, Dai H. Noncovalent functionalization of carbon nanotubes by fluorescein-polyethylene glycol: supramolecular conjugates with pH-dependent absorbance and fluorescence. Journal of the American Chemical Society 2007;129(9):2448–9.
- [26] Streu C, Meggers E. Ruthenium-induced allylcarbamate cleavage in living cells. Angewandte Chemie International Edition 2006;45(34):5645–8.
- [27] Xiang Y, Tong A, Jin P, Ju Y. New fluorescent rhodamine hydrazone chemosensor for Cu(II) with high selectivity and sensitivity. Organic Letters 2006;8 (13):2863-6.
- [28] Zhang X, Shiraishi Y, Hirai T. Cu(II)-selective green fluorescence of a rhodamine- diacetic acid conjugate. Organic Letters 2007;9(24):5039–42.

- [29] Lee MH, Kim HJ, Yoon S, Park N, Kim JS. Metal ion induced FRET off-on in tren/ dansyl-appended rhodamine. Organic Letters 2008;10(2):213-6.
- [30] Yang XF, Liu P, Wang L, Zhao M. A chemosensing ensemble for the detection of cysteine based on the inner filter effect using a rhodamine B spirolactam. Journal of Fluorescence 2008;18(2):453–9.
- [31] Yang YK, Yook KJ, Tae J. A rhodamine-based fluorescent and colorimetric chemodosimeter for the rapid detection of Hg<sup>2+</sup> ions in aqueous media. Journal of the American Chemical Society 2005;127(48):16760-1.
- [32] Ko SK, Yang YK, Tae J, Shin I. In vivo monitoring of mercury ions using a rhodamine-based molecular probe. Journal of the American Chemical Society 2006:128(43):14150–5.
- [33] Yang H, Zhou Z, Huang K, Yu M, Li F, Yi T, et al. Multisignaling optical- electrochemical sensor for Hg<sup>2+</sup> based on a rhodamine derivative with a ferrocene unit. Organic Letters 2007;9(23):4729–32.
- [34] Soh JH, Swamy KMK, Kim SK, Kim S, Lee SH, Yoon J. Rhodamine urea derivatives as fluorescent chemosensors for Hg<sup>2+</sup>. Tetrahedron Letters 2007;48 (34):5966–9.
- [35] Zhang M, Gao Y, Li M, Yu M, Li F, Li L, et al. A selective turn-on fluorescent sensor for Fe<sup>III</sup> and application to bioimaging. Tetrahedron Letters 2007;48 (21):3709–12.
- [36] Chen X, Ma H. A selective fluorescence-on reaction of spiro form fluorescein hydrazide with Cu(II). Analytica Chimica Acta 2006;575(2):217–22.
- [37] Yang XF, Li Y, Bai Q. A highly selective and sensitive fluorescein-based chemodosimeter for Hg<sup>2+</sup> ions in aqueous media. Analytica Chimica Acta 2007;584(1):95–100.
- [38] Burdette SC, Frederickson CJ, Bu W, Lippard SJ. ZP4, an improved neuronal Zn<sup>2+</sup> sensor of the zinpyr family. Journal of the American Chemical Society 2003;125(7):1778–87.
- [39] Nolan EM, Burdette SC, Harvey JH, Hilderbrand SA, Lippard SJ. Synthesis and characterization of zinc sensors based on a monosubstituted fluorescein platform. Inorganic Chemistry 2004;43(8):2624–35.
- [40] Chang CJ, Nolan EM, Jaworski J, Okamoto KI, Hayashi Y, Sheng M, et al. ZP8, a neuronal zinc sensor with improved dynamic range; imaging zinc in hippocampal slices with two-photon microscopy. Inorganic Chemistry 2004;43(21):6774—9.
- [41] Nolan EM, Racine ME, Lippard SJ. Selective Hg(II) detection in aqueous solution with thiol derivatized fluoresceins. Inorganic Chemistry 2006;45 (6):2742–9
- [42] Nolan EM, Jaworski J, Racine ME, Sheng M, Lippard SJ. Midrange affinity fluorescent Zn(II) sensors of the zinpyr family: syntheses, characterization, and biological imaging applications. Inorganic Chemistry 2006;45(24): 9748-57.
- [43] Jun EJ, Kim JA, Swamy KMK, Park S, Yoon J. A fluorescein derivative for nanomolar aqueous copper and monitoring copper ion uptake by transferrin and amyloid precursor protein. Tetrahedron Letters 2006;47(7): 1051-4.
- [44] Kim MH, Noh JH, Kim S, Ahn S, Chang SK. The synthesis of crown etherappended dichlorofluoresceins and their selective Cu<sup>2+</sup> chemosensing. Dyes and Pigments 2009;82(3):341–6.
- [45] Meng XM, Zhu MZ, Liu L, Guo QX. Novel highly selective fluorescent chemosensors for Zn(II). Tetrahedron Letters 2006;47(10):1559–62.
- [46] Meng XM, Liu L, Hu HY, Zhu MZ, Wang MX, Shi J, et al. Highly sensitive and selective fluorescent chemosensors for Hg(II) in an aqueous environment based on carbamodithioate. Tetrahedron Letters 2006;47(45):7961–4.
- [47] Yang ZY, Yang RD, Li FS, Yu KB. Crystal structure and antitumor activity of some rare earth metal complexes with Schiff base. Polyhedron 2000;19 (26–27):2599–604.
- [48] Jia Q, Liao W, Li D, Niu C. Synergistic extraction of lanthanum(III) from chloride medium by mixtures of 1-phenyl-3-methyl-4-benzoyl-pyrazalone-5 and triisobutylphosphine sulphide. Analytica Chimica Acta 2003;477 (2):251–6.
- [49] Wang Y, Yang ZY. Crystal structure of Ni(II) complex and fluorescence properties of Zn(II) complex with the Schiff base derived from diethenetriamine and PMBP. Journal of Luminescence 2008;128(3):373–6.
- [50] Benesi HA, Hildebrand JH. A spectrophotometric investigation of the interaction of iodine with aromatic hydrocarbons. Journal of the American Chemical Society 1949;71(8):2703-7.
- [51] Barra M, Bohne C, Scaiano JC. Effect of cyclodextrin complexation on the photochemistry of xanthone. Absolute measurement of the kinetics for triplet-state exit. Journal of the American Chemical Society 1990;112 (22):8075–9.
- [52] Margulies D, Melman G, Shanzer A. A molecular full-adder and full-subtractor, an additional step toward a moleculator. Journal of the American Chemical Society 2006;128(14):4865–71.
- [53] Tsuge K, DeRosa F, Lim MD, Ford PC. Intramolecular reductive nitrosylation: reaction of nitric oxide and a copper(II) complex of a cyclam derivative with pendant luminescent chromophores. Journal of the American Chemical Society 2004;126(21):6564–5.
- [54] Lim MH, Lippard SJ. Metal-based turn-on fluorescent probes for sensing nitric oxide. Accounts of Chemical Research 2007;40(1):41–51.